
Note

**3-AMINO-3-DEOXYGLUCOSE
PRODUCED BY A
STREPTOMYCES SP.**

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We have discovered that 3-amino-3-deoxyglucose is produced by a new *Streptomyces* species plated out from a Kansas soil. This antibiotic is a constituent of kanamycin¹⁾ which was produced by *Streptomyces kanamyceticus* and of 3-trehalosamine²⁾ which was produced by *Nocardiosis trehalosei*. This constitutes the first report of the production of 3-amino-3-deoxyglucose by an organism other than a *Bacillus*^{3,4)}. This antibiotic was shown to inhibit cell wall synthesis⁴⁾. Two syntheses have been reported^{5,6)}.

The organism was characterized and named *Streptomyces lansus* DIETZ and LI sp. nov. (UC[®] 8204). L-Diaminopimelic acid was found in the whole cell hydrolysate⁷⁾. This organism displayed many similarities to *Streptomyces venetus*⁸⁾ DS 24288 (NRRL 3987, UC[®] 5789) on Ektachrome⁹⁾. Both are melanin-positive and have spiral spore chains bearing spiny to hairy spores. They are distinguished from one another in their growth on carbon compounds in synthetic medium¹⁰⁾ and by the fact that *S. lansus* grows well at 45°C while *S. venetus* does not grow at 45°C. Antibiotic production and other general cultural and biochemical differences also distinguish the two organisms.

A culture of *S. lansus* on HICKNEY-TRESNER agar was used to inoculate eight 500-ml flasks which contained 100 ml of sterile growth medium each. The medium consisted of 5 g cerelose, 30 ml Brer Rabbit molasses, 10 g sodium glutamate, 2 g ammonium sulfate and 2 g sodium chloride per liter of tap water and was adjusted to pH 7.2 with 5 N sodium hydroxide solution prior to sterilization. The flasks were shaken on a rotary shaker (250 rpm) for three days at 28°C.

The contents were used to inoculate 10 liters of the same medium in a fermentation jar. At 28°C the maximum antibiotic concentration was achieved on the 5th day as determined by antibacterial activity on agar trays seeded with *Bacillus subtilis* UC[®] 564.

Nine liters of beer grown as above was filtered at harvest pH over a pad of Dicalite 4200. The filtrate was adjusted to pH 7 and percolated over a 50×3.8 cm column of granular charcoal. The charcoal bed was washed with deionized water until the eluate was clear. The antibiotic was eluted with 1:1 acetone-water (v/v). The acetone was removed on a rotary evaporator from the active fraction and the aqueous solution was passed over a 50×3.8 cm bed of Dowex 50W×8 (H⁺). The resin was washed with deionized water and the antibiotic was eluted with a water to 1.0 M ammonium sulfate gradient. The active fractions were pooled and desalted on a charcoal column as above. Removal of the acetone followed by lyophilization of the aqueous, desalted solution yielded 3.0 g of slightly yellow solid.

The product was reducible with aniline phthalate solution (0.93 g aniline and 1.66 g phthalic acid in 100 ml *n*-butanol saturated with water). The 100 MHz PMR spectrum in D₂O at pD 3 with external TMS showed anomeric doublets at δ 5.45 ($J=3.5$ cps) and δ 4.90 ($J=7.0$ cps). This suggested a mixture of anomers of a monosaccharide.¹¹⁾ The CMR spectrum in D₂O at pD 3 with external TMS showed pairs of peaks as follows: δ 97.4 and 92.6 (D), δ 78.1 and 72.4 (D), δ 72.7 and 69.7 (D), δ 67.9 and 67.6 (D), δ 61.7 and 61.5 (T), δ 59.2 and 56.3 (D). This spectrum was nearly identical to that of an authentic mixture of 3-amino-3-deoxy- α - and β -D-glucose obtained from *Bacillus aminoglucosidicus*³⁾ kindly provided to us by Dr. B. J. MAGERLEIN of The Upjohn Company.

A sample was derivatized with trimethylsilyl imidazole followed by trifluoroacetic anhydride. The per-O-trimethylsilyl-N-trifluoroacetyl derivative was analyzed on a Hewlett-Packard Model 5992 desktop GC-MS spectrometer. The column (0.4×80 cm) was packed with 3% OV-17 on high efficiency Chromosorb W. Peaks were observed at 9.7 and 10.8 minutes using a thermal gradient

of 3°C/min starting at 140°C. The mass spectra and retention times of these peaks were identical to those obtained for the derivatized, authentic α , β mixture of 3-amino-3-deoxy-D-glucose.

Methanolysis of a second sample followed by derivatization and analysis as described above gave a mixture of the protected methyl α , β -glycosides. The retention times and fragmentation patterns of the major peaks were identical to those obtained for the comparable derivative of the basic sugar from 3-trehalosamine isolated from *Nocardiosis trehalosei*²⁾.

Thus, 3-amino-3-deoxy-D-glucose is one of a very few antibiotics that are produced both by a unicellular bacterium (family *Bacillaceae*) and a filamentous bacterium (family *Streptomycetaceae*). It is the only amino sugar antibiotic discovered thus far that is produced by both groups of microorganisms.

References

- 1) UMEZAWA, H.; K. MAEDA, M. UEDA, Y. YAGISHITA, S. KONDO, Y. OKAMI, R. UTAHARA, Y. OSATO, K. NITTA & T. TAKEUCHI: Production and isolation of a new antibiotic kanamycin. *J. Antibiotics, Ser. A* 10: 181~188, 1957
- 2) DOLAK, L. A.; T. M. CASTLE & A. L. LABORDE: 3-Trehalosamine, a new disaccharide antibiotic. *J. Antibiotics* 33: 690-694; 1980
- 3) UMEZAWA, S.; K. UMINO, S. SHIBAHARA, M. HAMADA & S. OMOTO: Fermentation of 3-amino-3-deoxy-D-glucose. *J. Antibiotics, Ser. A* 20: 355~360, 1967
- 4) IWAI, Y.; H. TANAKA, R. ŌIWA, S. SHIMIZU & S. ŌMURA: Studies on bacterial cell wall inhibitors. III. 3-Amino-3-deoxy-D-glucose, an inhibitor of bacterial cell wall synthesis. *Biochim. Biophys. Acta* 498: 223~228, 1977
- 5) BAER, H. H.: Synthetic kanosamine. *J. Am. Chem. Soc.* 83: 1882~1885, 1961
- 6) GUTHRIE, R. D.: The reaction of periodate-oxidized methyl 4, 6-O-benzylidene- α -D-glucopyranoside with phenylhydrazine: A new synthesis of methyl 3-amino-3-deoxy- α -D-glucopyranoside derivatives. *Proc. Chem. Soc.* 1960: 387~388, 1960
- 7) BECKER, B.; M. P. LECHEVALIER & H. A. LECHEVALIER: Chemical composition of cell wall preparations from strains of various form-genera of aerobic actinomycetes. *Appl. Microbiol.* 13: 236~243, 1966
- 8) BELLOC, A.; J. FLORENT, J. LUNEL, D. MANCY & J. VERNER: Thrombolytic and fibrinolytic enzyme from *Streptomyces*. U.S. Patent 3,875, 005, 1975
- 9) DIETZ, A.: Ektachrome transparencies as aids in actinomycete classification. *Ann. N.Y. Acad. Sci.* 60: 152~154, 1954
- 10) SHIRLING, E. B. & D. GOTTLIEB: Methods for characterization of *Streptomyces* species. *Int. J. Syst. Bacteriol.* 16: 313~340, 1966
- 11) KAMERLING, J. P.; M. J. A. DEBIE & J. F. G. Vliegenthart: A PMR study of the anomeric protons in pertrimethylsilyl oligosaccharides, a determination of the configuration of the glycosidic bond. *Tetrahedron* 28: 3037~3047, 1972
- 12) BUCHANAN, R.E. & N.E. GIBBONS (ed.): *BERGEY'S Manual of Determinative Bacteriology*, 8th ed. The Williams & Wilkins Co., Baltimore, 1974